

NMR Facility

Introductory Lecture

Dr. Nick Rees

Dr. Harry Mackenzie

Outline

NMR Facility Staff

Magnet Hazards and Safety

Sample Preparation

Data Processing

Facilities and Instrumentation

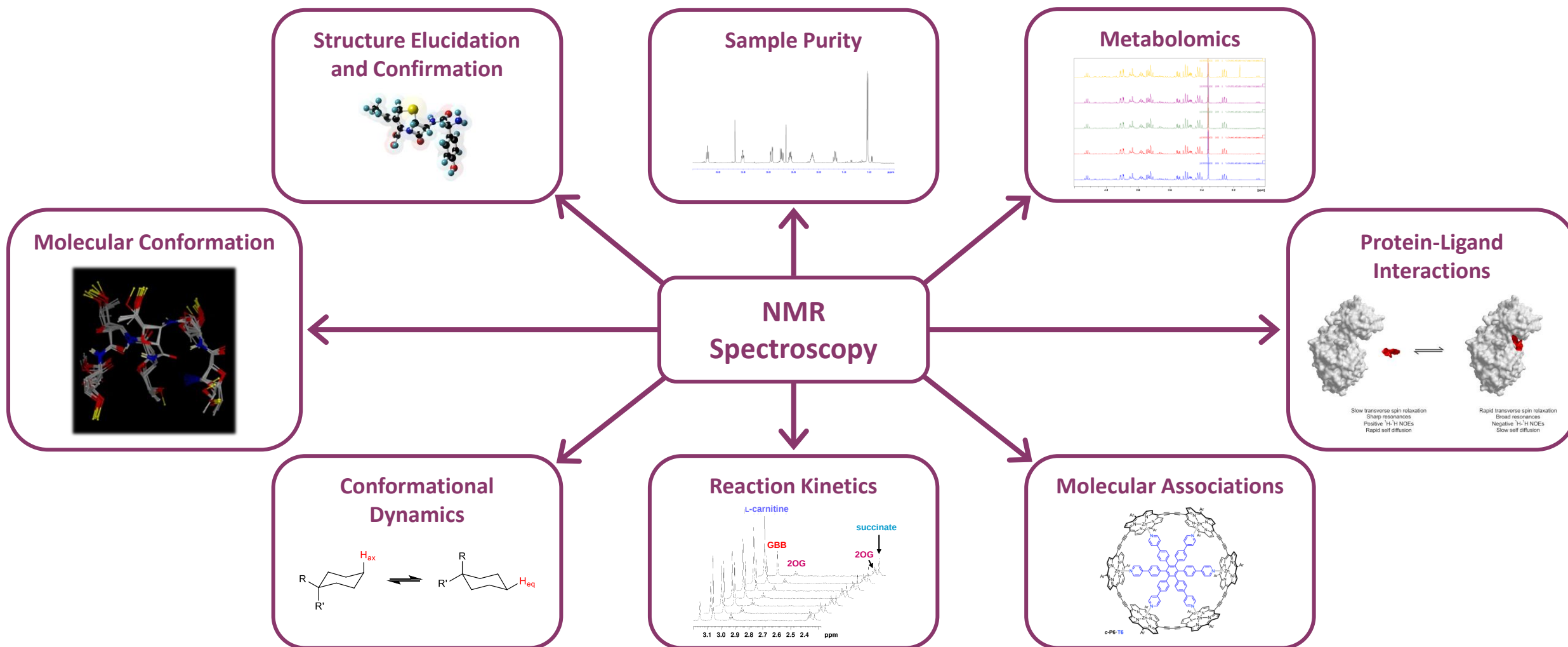
Open Access Facilities

NMR Submission Service

Online Resources: NMR web site

Future Training Courses





Facility Staff

Dr. Nick Rees

Head of Inorganic and Solid-State NMR

Dr. Harry Mackenzie

Head of Organic and Biological NMR

Dr. Coral Mycroft

Service Manager

~~Caitlin Salter~~ Edward Tomlinson (Maternity)

Service Technician



Nick



Harry



Coral



Edward

EMAIL US!



nmrstaff@maillist.chem.ox.ac.uk

Safety in the NMR Laboratories

Very Strong Magnetic Fields!

Hazards to:

heart pacemakers

magnetic bank, ID cards

watches (non-LCD)

Stray fields in corridors!

especially ground floor NMR

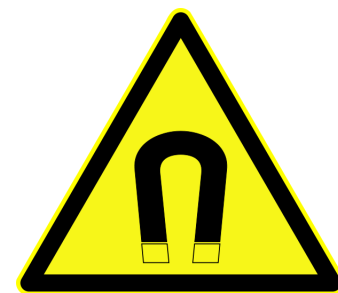


Cryogenic Liquids: extreme cold, oxygen depletion

Safety Rules

Clean Spaces: No laboratory coats or gloves in NMR labs.

No metal objects to be taken into NMR labs.



Sample breakages must be dealt with **immediately**

Inform the NMR staff in all cases: nmrstaff@maillist.chem.ox.ac.uk



Do not enter labs during weekly liquid nitrogen re-fills

Accessibility in the NMR Laboratories

If you require any assistance or adjustments in relation to training and/or using this facility or if you have any concerns you would like to discuss beforehand, contact

nick.rees@chem.ox.ac.uk

or

harry.mackenzie@chem.ox.ac.uk

Sample Preparation

Tubes and *deuterated* solvents from stores

Tubes must be “Wilmad 507” or “Norell S400” grade (or equivalent) at least

Tubes must not be scratched or broken

Label tubes very carefully (**Group & User Initials minimum**)

Solutions must be correct depth (4 cm)

Solutions must be free from particulates

Sample Preparation: Bad Practice



NMR Cap Colour and Tube Labelling Rules

The following rules are in place for the safety of all users of the NMR facility and must be followed. This includes NMR tubes used for open-access, hands-on, and the submission service.

Tube caps may be ordered from VWR on R12 as bags of 100.

Rules:

All groups must use their **group cap colour**

The top of the cap must be labelled with the **surname initial of your PI**

Your own **name/initials** must be labelled on the tube

The **solvent** used must be labelled on the tube

NOTE: If you are using a J Young's tube, you may use any colour cap but the full group initials must be written on, as well as the other information

Cap Colours: Organic and Chemical Biology

A E. Anderson

A H. Anderson

A A. Avestro

A Y. Aye

B H. Bayley

B J. Burton

C D. Congrave

D D. Dixon

A T. Donohoe

F S. Fletcher

F M. Fuchter

G V. Gouverneur

M I. McCulloch

M P. McGonigal

Q Y. Qing

R P. Rabe

R J. Robertson

R A. Russell

S C. Schofield

S M. Smith

W M. Willis

Cap Colours: Inorganic Chemistry

A	S. Aldridge	M	M. Mehta	
B	P. Beer	N	M. Neidig	
D	J. Davis	O	D. O'Hare	
F	S. Faulkner	T	E. Tsang	
G	G. Gregory	V	K. Vincent	
H	R. Hoye	W	C. Williams	N NMR
L	M. Langton	W	L. Wong	X External

Sample Masses Required

Rule of thumb for high-quality spectra

400 MHz open-access spectrometers:

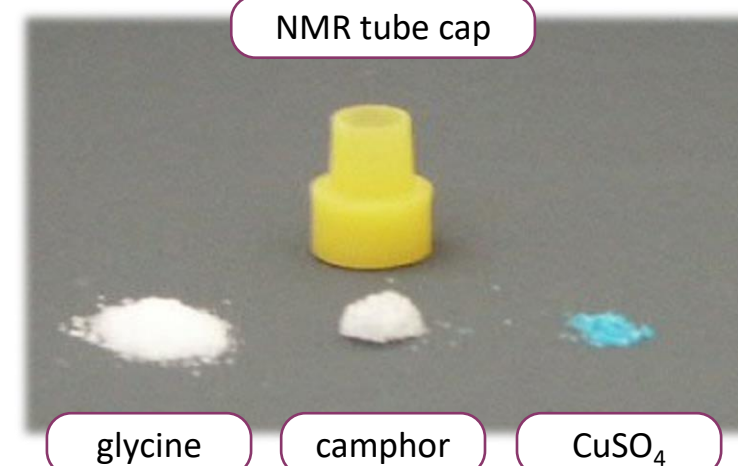
*Proton & 2D COSY: **2 mg***

*2D H-C HSQC: **10 mg***

*1D Carbon: **20 mg***

Please weigh your NMR samples!!

How much is 10 mg?



Data Processing

Data from all spectrometers can **ONLY** be downloaded from the SRF server for offline processing and local storage

Windows:

\\chem.ox.ac.uk\srf\nmr

macOS:

smb://chem.ox.ac.uk/srf/nmr

PLEASE no USB drives

Instrument	Year	Group	Sample
 AV600	 2012	 ajrggrp	 AAM-668-A-crude
 AV700	 2013	 cjsgrp	 AAM-668-B-crude
 AVB400	 2014	 dcgrp	 AR-518 Crude
 AVB500	 2015	 djdgrp	 AR-523 D
 AVD500	 2016	 eaagrp	 AR-523 E
 AVF400	 2017	 hlagrp	 AR-523 F
 AVG400	 2018	 imggrp	 AR-524
 AVH400	 2019	 jwbgrp	 AR-525 Crude
 AVX500	 2020	 mcwgrp	 AR-602 Yield
 bgd_backup	 2021	 mdsgrp	 AR-646 Yield
 DPX200	 2022	 nmrstaff	 AR-679
 External_data	 2023	 ontgrp	 AR-680
 HFX400	 2024	 prgrp	 CB-5-23-19-23-Aug14-2025-30-
 HG400	 2025	 sjcgrp	 CB-5-26-ACIDIC-Apr30-2025-8-
 HXY400		 spfgrp	 CB-5-26-CRUDE-Apr30-2025-7-
 NEO600		 tbgrp	 CB-5-29 PC-Aug14-2025-29-
		 tjdgrp	 CB-5-38B-CRUDE-Jun05-2025-59-
		 tlhgrp	 CB-5-52A-CRUDE-Jul15-2025-33-
		 vggrp	

Data Processing Software

Windows and macOS:

MestreNova (MestreLab):

Platform independent, 1D and 2D processing

Chemistry site licence



Windows and macOS:

TopSpin (Bruker):

used on all spectrometers, 1D and 2D processing

free for academic use



Software Installation

MestreNova 16.0:

Download from the Mestrelab website: <https://mestrelab.com/download>

Drag and drop licence from NMR server: /NMR Software/Mnova/

TopSpin 4.5:

Download from Bruker website and request licence:

<https://store.bruker.com/en-GB/products/topspin-for-processing-academic-government/>

NMR Facility – 4 Levels Of Analysis

Open Access

Walk up use at any time of the day

Common 1D/2D experiments: ^1H , ^{13}C , ^{19}F , ^{31}P , DEPT, COSY, HSQC, HMBC, NOESY**

Fast turnover and hence rapid sample throughput

Ground/first floors: Organic Chemistry and Chemical Biology

Second floor: Inorganic Chemistry

Hands on

Users can book the basement systems for manual/automated operation

NMR Service

A submission service where researchers may submit samples for the NMR staff to run on the basement instruments.

Research Projects

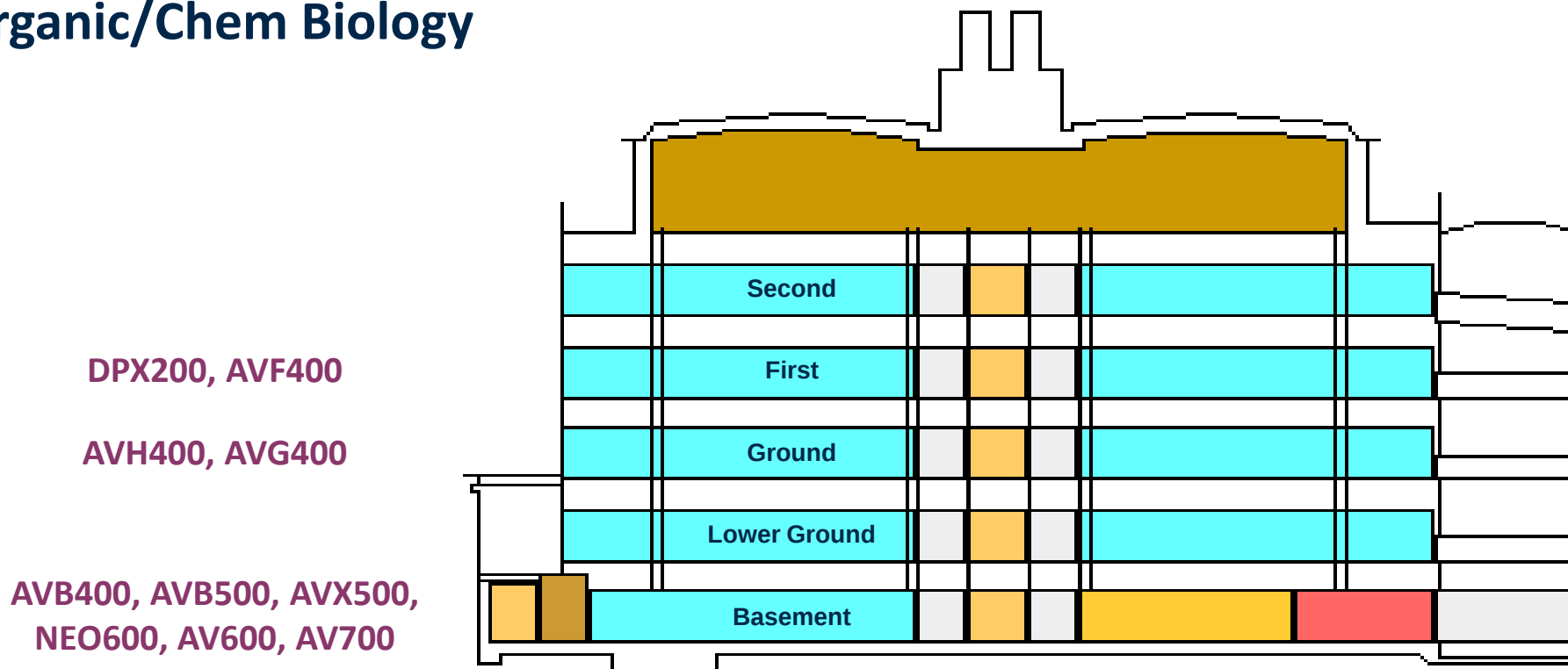
Collaborative projects involving the NMR staff/group

NMR in CRL

[Instrument nicknames shown]

Organic/Chem Biology

Inorganic Section



Venus400, Hg400

AVD500, HFX400, HXY400

Instrument Queues and Status

Open Access instrument queues(beta):

<https://intranet.chem.ox.ac.uk/nmrq/>



NMR Facility NMR Queues

GROUND FLOOR

AVH400

RUNNING

Day: 00:01 Night: 00:31



Updated 13:43:22 • Next refresh in ~60s

AVG400

RUNNING

Day: 00:04 Night: 00:58



Updated 13:43:22 • Next refresh in ~60s

FIRST FLOOR

DPX200

RUNNING (IDLE)

Day: 00:00 Night: 00:00



Updated 13:43:22 • Next refresh in ~60s

AVF400

RUNNING

Day: 00:04 Night: 01:03



Updated 13:43:22 • Next refresh in ~60s

SECOND FLOOR

HG400

RUNNING (IDLE)

Day: 00:00 Night: 00:00



Updated 13:43:22 • Next refresh in ~60s

Venus400

RUNNING (IDLE)

Day: 00:00 Night: 00:26



Updated 13:43:22 • Next refresh in ~60s

Legend

Running Completed Failed Load Failed ATM Failed LOCK Failed Shim Failed Acquisition Failed Other



IconNMR Automation Run Status:
Day Experiments: 00:08
Busy Until: Tue 14:10
Night Experiments: 00:38

Run	Queue	Name	Exp-Comment	Type	Host	Start	End	Time
1	Completed	Sep30-2025-SB74 RET	1 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	hmagp	00:01:32	Instrument		
2	Completed	HA15287 400G	1 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	jvrbgpp	00:01:32	Instrument		
3	Completed	Sep29-2025-3-LG-170	1 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	hmagp	00:01:32	Instrument		
4	Completed	KC-811A	1 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	ddgpp	00:01:32	Instrument		
4	Completed	KC-811A	2 N CRL_FLUORINE 19F (1H coupled, 0 to 250 ppm, 16 scans)	ddgpp	00:00:48	Instrument		
5	Completed	KC-811B	3 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	ddgpp	00:01:32	Instrument		
5	Completed	KC-811B	4 N CRL_FLUORINE 19F (1H coupled, 0 to 250 ppm, 16 scans)	ddgpp	00:00:48	Instrument		
6	Completed	CML-06-15RAC	1 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	mdgpp	00:01:32	Instrument		
7	Completed	Sep29-2025-7-ade905_c	1 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	ddgpp	00:01:32	Instrument		
8	Completed	DC-679-NOESY-Sep29-2025-S-	1 a CRL_NOESY 1H-1H NOESY	ddgpp	00:24:00	Instrument		
9	Completed	ALF118_prev	1 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	esgpp	00:01:32	Instrument		
11	Completed	GV-812 R2T3-12	1 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	vsgpp	00:01:32	Instrument		
11	Completed	GV-812 R2T3-12	2 N CRL_FLUORINE 19F (1H coupled, 0 to 250 ppm, 16 scans)	vsgpp	00:00:48	Instrument		
12	Completed	GV-812 R2T1-R2T2	1 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	vsgpp	00:01:32	Instrument		
12	Completed	GV-812 R2T1-R2T2	2 N CRL_FLUORINE 19F (1H coupled, 0 to 250 ppm, 16 scans)	vsgpp	00:00:48	Instrument		
13	Completed	IDH-014-090-2-A-CDCL3 Sep29-2025-13-	1 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	agpp	00:01:32	Instrument		
13	Completed	IDH-014-090-2-A-CDCL3 Sep29-2025-13-	2 N CRL_FLUORINE 19F (1H coupled, 0 to 250 ppm, 16 scans)	agpp	00:00:48	Instrument		
13	Completed	IDH-014-090-2-A-CDCL3 Sep29-2025-13-	3 N CRL_FLUORINE_HDEC 19F (1H decoupled, 0 to 250 ppm, 16 scans)	agpp	00:00:48	Instrument		
14	Completed	D1Wn363-363-V9	1 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	cspp	00:01:32	Instrument		
14	Completed	D1Wn363-363-V9	2 a CRL_COSY 1H-1H COSY	cspp	00:05:07	Instrument		
15	Completed	ALF118_dea2	1 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	esgpp	00:01:32	Instrument		
23	Completed	KC-811Bather	1 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	ddgpp	00:01:32	Instrument		
23	Completed	KC-811Bather	2 N CRL_FLUORINE 19F (1H coupled, 0 to 250 ppm, 16 scans)	ddgpp	00:00:48	Instrument		
24	Completed	HA15435 400G	2 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	jvrbgpp	00:01:32	Instrument		
25	Completed	KC-811ISM	1 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	ddgpp	00:01:32	Instrument		
25	Completed	KC-811ISM	2 N CRL_FLUORINE 19F (1H coupled, 0 to 250 ppm, 16 scans)	ddgpp	00:00:48	Instrument		
26	Completed	Sep30-2025-24-LG-360-2	1 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	hmagp	00:01:32	Instrument		
27	Completed	IDH-014-090-3-A-CDCL3 Sep29-2025-27-	1 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	agpp	00:01:32	Instrument		
28	Completed	AB-4253 spin 1-1 TEAF-U-rt	1 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	vsgpp	00:01:32	Instrument		
28	Completed	AB-4253 spin 1-1 TEAF-U-rt	2 N CRL_FLUORINE 19F (1H coupled, 0 to 250 ppm, 16 scans)	vsgpp	00:00:48	Instrument		
29	Completed	KC-811B-top	1 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	ddgpp	00:01:32	Instrument		
29	Completed	KC-811B-top	2 N CRL_FLUORINE 19F (1H coupled, 0 to 250 ppm, 16 scans)	ddgpp	00:00:48	Instrument		
30	Completed	KC-811B-mid	3 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	ddgpp	00:01:32	Instrument		
30	Completed	KC-811B-mid	4 N CRL_FLUORINE 19F (1H coupled, 0 to 250 ppm, 16 scans)	ddgpp	00:00:48	Instrument		
31	Completed	Sep30-2025-31-JPE2-F14	1 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	agpp	00:01:32	Instrument		
31	Completed	Sep30-2025-31-JPE2-F14	2 a CRL_CARBOON 13C (256 scans)	agpp	00:14:33	Instrument		
32	Completed	Sep30-2025-32-JPE2-F13	1 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	agpp	00:01:32	Instrument		
32	Completed	Sep30-2025-32-JPE2-F13	2 a CRL_CARBOON 13C (256 scans)	agpp	00:14:33	Instrument		
33	Completed	Sep30-2025-33-JPE2	1 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	agpp	00:01:32	Instrument		
33	Completed	Sep30-2025-33-JPE2	2 a CRL_CARBOON 13C (256 scans)	agpp	00:14:33	Instrument		
37	Completed	HA15435 400G	1 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	jvrbgpp	00:01:32	Instrument		
38	Completed	NC176_SMI	1 N CRL_PROTON 1H (-4 to 16 ppm, 16 scans)	vsgpp	00:01:32	Instrument		

History List	Run	Queue	Name	Exp-Comment	Type	Host	Start	End	Time
42	13:52:26	Sep30-2025-42-SM4-033	3 CRL_PHOSPHORICUS_HDEC	agpp	-	-	-	-	-
42	13:48:38	Sep30-2025-42-SM4-033	1 CRL_PROTON	agpp	-	-	-	-	-
43	13:48:48	Sep30-2025-41-SM4-028	3 CRL_PHOSPHORICUS_HDEC	agpp	-	-	-	-	-
43	13:41:55	Sep30-2025-41-SM4-028	1 CRL_PROTON	agpp	-	-	-	-	-
33	13:37:08	Sep30-2025-33-JPE2	1 CRL_PROTON	agpp	-	-	-	-	-
32	13:39:33	Sep30-2025-32-JPE2-F13	2 CRL_CARBOON	agpp	-	-	-	-	-
33	13:34:45	Sep30-2025-33-LG-160-2-ecsk	1 CRL_PROTON	hmagp	-	-	-	-	-
32	13:39:34	Sep30-2025-32-JPE2-F13	1 CRL_PROTON	agpp	-	-	-	-	-
31	13:06:06	Sep30-2025-31-JPE2-F14	1 CRL_PROTON	agpp	-	-	-	-	-
46	12:54:03	JR02771-TM2-2b	2 CRL_PROTON_QUANT	esgpp	-	-	-	-	-
45	12:48:55	JR02771-TM2-2b	1 CRL_PROTON_QUANT	esgpp	-	-	-	-	-
1	12:28:19	Sep30-2025-SB74 RET	1 CRL_PROTON	hmagp	-	-	-	-	-
25	12:23:16	KC-811ISM	2 CRL_FLUORINE	ddgpp	-	-	-	-	-
25	12:19:05	KC-811ISM	1 CRL_PROTON	ddgpp	-	-	-	-	-
24	12:14:45	HA15435 400G	2 CRL_PROTON	jvrbgpp	-	-	-	-	-
40	12:08:50	Sep30-2025-40-RBC_P1B1_4bk	1 CRL_PROTON	hmagp	-	-	-	-	-
39	12:03:21	NC176_SMI_2nd	1 CRL_PROTON	vsgpp	-	-	-	-	-

Organic Chemistry and Chemical Biology

10 NMR spectrometers available:

1 x 200 MHz: **Open Access** ^1H only

3 x 400 MHz: **Open Access** ^1H , ^{13}C , ^{19}F , ^{31}P , routine 2Ds

1 x 400 MHz: **Hands on**

2 x 500 MHz: **Hands on** and **Service**

1 x 600 MHz: **Service**

1 x 600 MHz: **Hands on**, **Service**, **Research Projects**

1 x 700 MHz: **Hands on**, **Service**, **Research Projects**

Organic Chemistry and Chemical Biology

Open Access

Ground Floor



AVG400



AVH400



AVF400



DPX200

High-Field Facilities



Basement high-field NMR lab

400 and 500 MHz instruments available for **hands on** use

600/700 MHz for bio-projects

Training **must** be given by NMR staff

Please enquire with NMR Staff if you wish to be trained

Online booking of instrument time: <https://booking.chem.ox.ac.uk/>

NMR Submission Service

Availability of experiments

Other nuclei: ^2H , ^7Li , ^{11}B , ^{27}Al , ^{119}Sn etc.

2D experiments: TOCSY, H2BC, ROESY, ^1H - ^{15}N HSQC/HMBC

More specialised experiments: DOSY, variable temperature, pure shift, selective and more

Dilute samples

Walk-up instruments do not allow you to adjust any experiment parameters

The NMR service will adjust experiment parameters so acceptable spectra is obtained.

Higher field spectrometers

500 – 700 MHz

Solid state samples



NMR Submission Service (New System)

NMR Sample Submission Service v.24 Nov 2023

Welcome [Edward Tomlinson \(NMR\)](#)

To submit a new sample, [fill in this online form](#)

List of your previous sample submissions and their status:

Rows: 0-0 / 0		Records: 25 ? Clear						
Sample Submission ID press on sample ID to add spectra	Your Ref. plus supplied supporting spectra	Sample Structure	Service Request	Submission Date	Cost Centre	NMR Service Time (hrs)	Instruments Used	Complete



New NMR Sample Submission

Sample for: **Mr Edward Tomlinson** (edward.tomlinson@chem.ox.ac.uk)

Date of Submission: **11/09/2025**

Group: **NMR Role: SRF Staff**

Phone: **75659**

Lab/Office:
CRL -00.086

Your Sample Ref Code/Number:

Project Cost Centre Code:

[Other - add into description](#)

Experiments:

☐ 1H ☐ 13C ☐ 19F ☐ 31P ☐ COSY ☐ HSQC ☐ HMBC ☐ NOESY ☐ Other (describe)

Indicate Hazards:

(select multiple if required, CTRL click)

☐ none
☐ unknown
☐ toxic
☐ carcinogen

Mass Supplied (mg):

Solvent

Sample Location: ☐ Rack ☐ Fridge ☐ Request

Describe the experiments that you would like the NMR service to run for you:

Upload a picture of your expected sample chemical structure, drag and drop to this area works.

No file chosen

[Submit Sample Request](#)

Inorganic Chemistry

5 NMR spectrometers available:

1 x 400 MHz: **Open Access** multinuclear

1 x 400 MHz: **Open Access** multinuclear

1 x 500 MHz: **Hands on** & **Service** multinuclear and VT work

1 x 400 MHz: **Service Solid State** HXY

1 x 400 MHz: **Service Solid State** HFX, microimaging & diffusion

Access to 600 MHz: **Service** multinuclear

Inorganic Chemistry

Open Access

Hands-on

Second Floor



AVG400



AVH400

Basement



AVD500

Solid state NMR

- Service provided by Dr Nick Rees
- Stable samples provided as a powdered solid (c.a. 200mg)
- Unstable samples can be packed in glove box
- Consult Nick Rees **before** submitting samples
- nick.rees@chem.ox.ac.uk
- Submit samples via the sample submission service
- Stable samples should be placed in the box through the basement NMR lab hatch
- For unstable samples provide email address on submission form.

2 x 400MHz Wide Bore 3 channel systems



HFX400

- 4 & 1.9 mm Triple Magic Angle Spinning Probes (^1H , ^{19}F , ^{31}P to ^{15}N)
- 4mm Low Gamma (^{13}C to ^{109}Ag) Magic Angle Spinning Probe
- Wideline Deuterium Probe
- Goniometer probe for oriented samples
- Variable temperature capable (-150 to 150C)



HFX400

- 3.2mm HFX Triple Magic Angle Spinning Probes (^1H , ^{19}F , ^{31}P to ^{15}N)
- 30mm Micro-imaging probe
- Diffusion probe (-150 to 200C)
- Variable temperature capable (-150 to 150C)

Solid State NMR Service

Sample Submission Form: Word document

List experiments

And specify SSNMR

If the sample needs to
be packed in glove box
put down as request

-
- ☐ HG400
 - ☐ HXY400
 - ☐ NMR Software
 - ☐ NMR Submissions
 - ☐ NMR_Staff

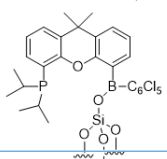


Solids Submission

INTERNAL: NMR SERVICE SUBMISSION FORM FOR DEPARTMENT OF CHEMISTRY

CHEMISTRY RESEARCH LABORATORY
NMR SERVICE

Name: Aisling Roper Status: D.Phil. Carousel #: CRL Lab: S12
Email: aisling.ropер@chem.ox.ac.uk Instr: Phone:
Group: Aldridge [Pt II's only] Lab. Supervisor's Name:
Submission Number:² 70957 Project Code/ Charge Account:³ DHT00110 DHSA.08 Submission Date: 06/07/23

Nuclei of interest: 31P, 11B **Sample @:¹ S2 glovebox – requires packing**
Structure:  Toxicity:⁵ unknown Solution Depths:⁹
Max 4.5 cm
Min 4.0 cm
Experiments required (list ALL): solid state 31P and 11B NMR coupled and decoupled
Nature of problem:⁶ expected 11B shift around +10-+60 ppm, expected 31P at approx. -10 ppm
Mass supplied:⁷ Solvent:⁸

Referencing: ¹H and ¹³C spectra are referenced externally to TMS in CDCl₃. ¹⁹F spectra and ³¹P spectra are referenced externally to CFCl₃ in CDCl₃, and to phosphoric acid in D₂O respectively. Indicate if you have added an internal reference.

- Part II students must provide the name of their laboratory supervisor.
- You should quote the number generated on the sample submission system page.
- This is what you would use for iProcurement purchases. Seek advice from the Finance team if you do not know this.
- Indicate where your sample can be found. If 'request', you will be contacted by the NMR staff in due course.
- Give ANY details you may know that relate to possible hazards associated with handling of the sample (such as in the case of sample spillage or tube breakages). E.g. toxic, carcinogen etc. If this is uncertain, enter UNKNOWN.
- Indicate the expected presence of unusual shifts. Describe briefly any particular problem you wish to address (this will help us choose the most appropriate experiment(s) for the problem). **All experiments requested must be listed on this form.**
- ¹H: 1-10 mg for the 600; ¹³C: 10+ mg for the 600 (ca. 50+ mg should be run on the 400s); ¹⁹F: 1-10mg; ³¹P: 10 mg. Please ask for others.
- For routine analysis, all samples should be supplied in 5 mm high-quality tubes (Norell 400S, Wilmad 507-PP, or New Era MP5 at least). Cracked, scratched or broken tubes will not be accepted.
- The maximum solvent depth for 5 mm tubes should be 4.5 cm (600µl), the minimum is 4.0 cm (500µl). Note that the automated spectrometers also require a sample depth of 4.0 - 4.5 cm. Samples with depths outside this range may be rejected.

Future Training Courses

Use of the Open-Access NMR Spectrometers & Service

Running this week and next: meet in CRL reception

Compulsory sessions: you must attend before using instruments or the NMR submission service. Thurs 2nd (pm), Mon 6th (pm), Tues 7th (am & pm).

MestreNova NMR Software Introductory Lecture

Single on-line lecture introducing main software features

Modern NMR Spectroscopy for the Research Chemist

8-lecture course providing overview of NMR techniques

This course can be found on Canvas at: <https://canvas.ox.ac.uk/courses/54457>

NMR Training for New Researchers

Organic/Chemical Biology Sections	Inorganic Sections
Register as a new user: New Organic/Chembio NMR User	Register as a new user: New Inorganic NMR User
Book a training session: Organic NMR Training	Book a training session: Inorganic NMR Training

Meet for training in CRL reception

To arrange training external users should email: nmrstaff@maillist.chem.ox.ac.uk

All the above links can be found in these slides online at: <https://nmr.chem.ox.ac.uk/>

QR for Training Sign-up Sheets

