ZipTip Protocol

Guidelines for Selecting ZipTip_{C18} or ZipTip_{C4} Pipette Tips



Solutions Required

Buffer A

100% Acetonitrile HPLC-grade

<u>Buffer B</u>

98% Milli-Q water
2% Acetonitrile HPLC-grade
0.1% Formic Acid (FA) for nanoLC or 0.1% Trifluoroacetic Acid (TFA) for MALDI

Buffer C

50% ACN 50% Milli-Q water 0.1% FA

Buffer D

Matrix CHCA or 2.5-DHB (20 mg/mL matrix in 30:70 (v/v) Acetonitrile: TFA 0.1% in water) - peptides

Matrix 2.5-DHAP (7.6 mg matrix in 375 μ l EtOH. Add 125 μ l of a solution containing 18mg/ml DAC dissolved in water) - Large Proteins

Matrix SA (20 mg/mL matrix in 30:70 (v/v) Acetonitrile: TFA 0.1% in water) - Proteins

Procedure

Note: Resin bed provides backpressure, so set pipette to 10 µl, depress plunger to dead stop and slowly release.

1. Acidify sample (Vol 20-100 μ l) by adding TFA (MALDI) or FA (nanoLC) to 0.1 % final concentration.

2. ZipTip equilibration

- Aspirate 10 µl **Buffer A** into tip and dispense to waste. Repeat.
- Aspirate 10 µl **Buffer B** into tip and dispense to waste. Repeat.
- **3.** Bind and Wash the peptides/proteins
 - Bind peptides to ZipTip pipette tip by aspirating and dispensing 3-7 cycles (simple mixtures), up to 10 cycles (complex).
 - Aspirate 10 µl **Buffer B** and dispense to waste. Repeat wash once more.

4. Elution:

nanoLC-MS	MALDI-TOF/TOF
Dispense 10µl of Buffer C into a clean Eppendorf tube. Aspirate and dispense elution solution through ZipTip at least 5 times without introducing air. Dispense 10µl of Buffer A into a clean Eppendorf tube	Dispense 1-3µl of Buffer D into a clean Eppendorf tube. Aspirate and dispense elution solution through ZipTip at least 5 times without introducing air.
Aspirate and dispense elution solution through ZipTip at least 5 times without introducing air. Combined the elution solutions into an empty Eppendorf tube.	
Dry in vacuum centrifuge.	
Resuspend in 10 µl Buffer B	
Transfer to a recovery vial	After 5 elution times through ZipTip spot on MALDI plate. Leave to dry